# Applications of Diatoms as Potential Microalgae in Nanobiotechnology

Ali Akbar Jamali<sup>1</sup>, Fariba Akbari<sup>2</sup>, Mohamad Moradi Ghorakhlu<sup>3</sup>, Miguel de la Guardia<sup>4</sup>, Ahmad Yari Khosroushahi<sup>2,5\*</sup>

<sup>1</sup>Department of Bioinformatics, Research Institute of Physiology and Biotechnology (RIPB), Zanjan University, Zanjan, Iran

<sup>2</sup>Research Center for Pharmaceutical Nanotechnology, Tabriz University of Medical Sciences, Tabriz, Iran

<sup>3</sup>Department of Biology, Faculty of Science, Zanjan University, Zanjan, Iran

<sup>4</sup>Department of Analytical Chemistry, University of Valencia, Dr. Moliner 50, 46100, Burjassot, Valencia, Spain

<sup>5</sup>Department of Pharmacognosy, Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran

ARTICLEINFO	ABSTRACT			
<i>Article Type:</i> Review Article	<i>Introduction:</i> Diatoms are single cell eukaryotic microalgae, which present in nearly every water habitat make them ideal tools for a wide range of applications such as oil explora-			
Article History: Received: 04 April 2012 Revised: 30 April 2012 Accepted: 02 May 2012 ePublished: 12 May 2012	tion, forensic examination, environmental indication, biosilica pattern generation, toxicity testing and eutrophication of aqueous ecosystems. <i>Methods:</i> Essential information on diatoms were reviewed and discussed towards impacts of diatoms on biosynthesis and bioremediation. <i>Results:</i> In this review, we present the recent progress in this century on the application of diatoms in waste degradation, synthesis of biomaterial, biomineraliza-			
<i>Keywords:</i> Diatoms Bioindicators Biosynthesis Bioremediation Biomineralization	tion, toxicity and toxic effects of mineral elements evaluations. <i>Conclusion:</i> Diatoms can be considered as metal toxicity bioindicators and they can be applied for biomineralization, synthesis of biomaterials, and degradation of wastes.			

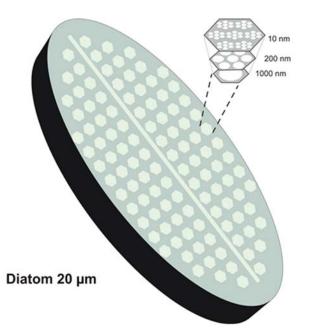
### Introduction

Diatoms are single cell eukaryotic microalgae, which are found in every habitat where water is present. Their abundance and wide distribution make them ideal tools for a wide range of applications both, as fossils and living organisms, (Atazadeh and Sharifi 2010). Examples of their wide range of applications include oil exploration, forensic examination and environmental indication. Today, the major emphasis are focused on their application in nanotechnology and biotechnology including nanofabrication techniques, chemo and biosensing, particle sorting, and control of particles in micro- and nano fluidics and also on their use in analyzing ecological problems; such as climate change, acidification, and eutrophication of aqueous ecosystems (Atazadeh and Sharifi 2010, Atazadeh et al 2007, Dolatabadi and de la Guardia 2011). Diatoms are highly robust organisms, and can inhabit virtually all photic zones from the equator to seemingly inhospitable sea ice where they are highly useful indicators of environmental conditions in their rapid response to environmental changes, including their capacity to react to sea ice

freezing around them with their "natural antifreeze" icebinding proteins (Chen *et al* 2010). Thus, in all climate zones, diatoms show an exceedingly high degree of flexibility that offer exciting possibilities in biotechnological applications despite challenging conditions.

The high degree of complexity and hierarchical structure displayed by diatom silica walls is achieved under mild physiological conditions. The biological processes that generate patterned biosilica are therefore of interest to the emerging field of nanotechnology. Biosilica and silicon, in their various forms, finds widespread use in electronic, optical, and structural materials. Research on uses of silicon and silica has been intense for decades, raising the question of how much diversity is left for innovation with this element (Dolatabadi and de la Guardia 2011, Losic et al 2009). Silica nanoparticles have proven to be important for several biotechnological and biomedical applications, such as biosensor design, drug delivery, cell labeling, cell separation, contrast agents for magnetic resonance and ultrasound medical imaging, and as a targeting and therapeutic platform for drug- or enzyme-released systems (Dolatabadi and de la

\*Corresponding author: Ahmad Yari Khosroushahi (PhD), Tel.: +98 (411) 3367914, Fax: +98 (411) 3367921, E-mail: yarikhosroushahia@tbzmed.ac.ir Copyright © 2012 by Tabriz University of Medical Sciences Guardia 2011). Silica bodies are opal phytoliths (phyto means 'plant' and lithos means 'rock' in Greek) produced by plants when soluble silica from the ground water is absorbed by the roots and carried to different parts of the plant system through the vascular system. Some plants have nonsiliceous phytoliths, such as calcium oxalate. Furthermore, the diatoms frustules could be used as complex templates in the patterning of biomolecules both at the micro and the nanoscale due to their intricate structural geometries (Yuan *et al* 2006). Figure 1 shows the structure of diatoms and their frustule holes in different scales which owes different features.



**Fig.1.** Schematic of the hierarchical structure of diatoms, showing different levels of structure from the nanoscale to the micrometer scales.

Diatoms have been used for toxicity testing (Atazadeh et al 2009, Florence and Stauber 1986). However, information obtained from such tests is based on the results for a few freshwater green algae that are easy to manipulate in culture (Mohan and Hosetti 1999). Although toxicity tests with isolated species can provide useful indications for environmental risk assessments of test compounds, they cannot predict changes at different organizational levels in natural communities (Berard et al 2002). Assessment of impacts of chemical contamination on the environment should take into account the natural variability of biological systems in space and time, and it is particularly noteworthy that any endpoint used to evaluate toxicity may be expected to vary in magnitude under different environmental and biological factors (Schindler 1987).

The taxonomic composition of benthic diatom communities has been widely used for monitoring water quality (Kelly 1998). Water quality monitoring programs have focused on comparisons of water chemistry with criteria derived ultimately from bioassays. However, assessment of community composition has several advantages over physical and chemical measurements of water quality and the use of biological communities as indicators of water quality is evolving as our understanding of the interactions between water quality and the integrity of biological communities improves (Atazadeh *et al* 2007).

In this paper, we try to show the feasibility of diatom frustules usage in biomineralization, synthesis of biomaterial, degrading waste and metal toxicity. Additionally, they have been used as bioindicators of water pollution in aquatic ecosystems.

#### Application of diatoms in biomineralization

Biomineralization is the process of producing mineral elements from organic compounds which is involved with biosystems. This process consists of methods, devices and requirements which could be supplied by microorganisms. To get this goal, bacteria, fungus, and diatoms are used largely. Diatoms due to their big plentiful are more commonly used than other organisms. In this case, diatoms can manufacture most of mineral materials. Biosilica, which is participate in diatoms structure, are being produced by diatoms, formed only in natural water and in the presence of sunlight without the need for high temperature or high-pressure treatment. Because of their unique structure, frustules find wide industrial application; they are used in water filters, building materials, and chromatography supports (Brunner et al 2009, Gutu et al 2009, Heredia et al 2008, Hildebrand 2008, Jeffryes et al 2008).

The production of platinum metal (Pt) has been recently showed by means of diatoms such as *Melosira nummuloides* in culture condition. In that study, the cultivation of *Melosira nummuloides* was demonstrated in the presence of dihydrogen hexachloroplatinate, hexahydrate in order to introduce Pt into frustules. The results indicated that Pt can be injected into frustules at the time of cultivation of living diatom cells (Yamazaki *et al* 2010).

Cadmium metal (Cd) is another element which can be produced by diatoms during their growth (Gutu *et al* 2009). Addition of cadmium sulphide to *Pinnularia sp.* Culture resulted in the appearance of Cd crystals in diatom's frustule. The results showed a simple and inexpensive chemical deposition process which can be utilized to coat intricately patterned diatom frustules biosilica with optoelectronic semiconductor thin films (Gutu *et al* 2009).

#### Application of diatoms in synthesis of biomaterial

Diatoms due to mentioned features and qualities are considered as unique machines in biosynthesis of biomaterial. Diatoms are ubiquitous and constitute an important group of the phytoplankton community having a major contribution to the total marine primary production. These products are being extracted and used in many cases. On the other hand, the more recent development of synthetic nanotechnology has been driven by the nearly insatiable global demand for ever smaller structures for electronic, optical, chemical, or biomedical devices (Bertrand 2010, Gordon et al 2005). Diatoms are considered as major group of phytoplankton that account for approximately 40% of the ocean carbon fixation and the vast majority of biogenic silica production through the construction of their cell walls. These microorganisms are used in synthesis of enzymes, biofuels, biosilica, and mineral materials, (Bertrand 2010, Haynes et al 2007, Schroder et al 2007). Common methods of silica synthesis mostly require high temperature conditions and extreme pH which imposes high costs to producers. However, microorganisms can manufacture silica and this process is well described in the most recent literature on the molecular biology of enzymemediated silica formation in marine and freshwater organisms, and the development of strategies towards the application of biocatalytically formed silica ("biosilica") that may be advantageous compared to silica synthesized by chemical methods. The synthesis of spicules is a rapid process. Studies on freshwater organisms revealed a growth rate of 5 µm per hour. Therefore, these microorganisms must accumulate large amounts of silicic acid from the surrounding water, which is undersaturated with respect to silicon, a process that obviously requires a lot of energy (Schroder et al 2007).

Enormous study findings recently described the methods of carotenoid biosynthesis in diatoms; these carotenoids consist of b-carotene, violaxanthin (Vio), diadinoxanthin (Ddx) and diatoxanthin (Dtx) and the chlorophyllous pigments, i.e. chlorophyll a, c1, c2 (Bertrand 2010). The carotenoid amount can be modified according to the environment and density of these compounds can be found by spectrochemical and chemical analysis. Besides, the biosynthetic pathway used to synthesis carotenoids is now quite well established in higher plants whilst many information are missing in diatoms (Bertrand 2010).

### **Evaluation of toxicity**

Several mineral elements are essential for living organisms at very low concentrations, but at high concentrations most of them are toxic and have a direct and adverse influence on various physiological and biochemical processes. These elements belong to the category of essential micronutrients and participate in growth, metabolism and enzyme activities. In contrast, other elements are basically toxics and they cause many problems and difficulties in natural media for the normal growth of organisms. One of the most studied toxic effects of mineral elements on microorganisms is growth inhibition. The degree of growth inhibition can be affected in different ways, depending on the strain of microalgae and metal concentration added in solution (Hess 2010, Liu *et al* 2010, Navarro *et al* 2008).

#### Application of diatoms in waste degradation

Wastes are unwanted substances invariably produced during daily activities. Depending on their physical state, they are classified as solid and liquid wastes and gaseous emissions. Sanitary refuse is the process of collection, transportation and disposal of solid wastes in a systematic, economic and hygienic way. Industrial wastes are the pollutant delivered out of a particular industry. The quality and quantity of wastes depend on the nature of industry, raw materials used, manufacturing housekeeping and process. Environment pollution is defined as contamination of water or alteration of the physical, chemical or biological properties of natural water. Water or other sources are said to be polluted when they change their quality or composition either naturally or because of human activities, thus becoming unsuitable for domestic, agricultural, industrial, recreational uses and for the survival of wildlife. Environment pollutant can be defined as an agent affecting aesthetic, physical, chemical and biological quality and wholesomeness of water (Ashok 2009, Evans and Furlong 2003, Pepper and Gerba 2005, Srinivas 2008, Yamazaki et al 2010).

These materials should be managed and controlled. Some methods are used in all over the world to overcome this problem. These methods consist of degradation physically, chemically or biologically. Because of releasing materials, chemically and physically methods do not be considered as a suitable for getting rid of these pollutants. Therefore, biological methods and their unique features qualified them to use in biodegradation of wastes. Biological methods involve the transformation or mineralization of contaminants to less toxic, more mobile, or more toxic but less mobile, forms. The main advantages of these methods are their ability to destroy a wide range of organic compounds, their potential benefit to soil structure and fertility and their generally nontoxic, 'green' image. Biodegradation process involves using bacteria, algae, and diatoms. Diatoms due to their unique structure and biological aspects are used largely in waste degradation (Glazer and Hiroshi 2007, Marchetti and Cassar 2009, Saade and Bowler 2009, Srinivas 2008).

As diatoms respond quickly to environmental changes and reflect both, physical and chemical characteristics of the overlying water masses, they are particularly useful for paleoecological reconstructions. These frustules accumulate and are partially preserved in the ocean sediments (Gordon *et al* 2005, Marchetti and Cassar 2009, Saade and Bowler 2009).

Diatoms, because of their structural and physiological features, can be used in material degradation. For example, some diatoms are removing the waste in environment and pollutants. In Malaysia, the degradation of aquatic pollution in the Pinang River Basin through diatoms was investigated (Maznah and Mansor 2002). In this investigation, different species of diatoms on different samples which gathered from many stations was employed. The results showed that the diatom community structure and the specific sensitivity of certain diatom species could be related to the degree of water quality in Pinang River Basin. The abundance of certain diatom species could be used as biological indicators to measure impacts of river pollution (Maznah and Mansor 2002).

Some researcher has been reported that certain freshwater green algae (e.g. Chlorella vulgaris, Scenedesmus platydiscus, Scenedesmus quadricauda and Selenastrum capricornutum) are capable of uptaking and degrading polycyclic aromatic hydrocarbons (PAHs) (Keum et al 2006, Li et al 2006, Luan et al 2006, Seo et al 2006, Seo et al 2007). Laminaria joapnica, a famous macroalgae, is growing in the coastal waters along the east coast of China uptakes and degrades of two PAHs phenanthrene and pyrene (Wang and Zhao, 2007). This macroalgae grown under different phenanthrene and pyrene concentrations showed variable stress responses as determined by their enzymatic activities (Wang and Zhao, 2007). This study finding showed that the Laminaria joapnica has a strong capacity to tolerate and uptake PAHs from seawater. Some recent studies have also reported that phenanthrene and pyrene could be transformed by micro-organisms into soluble diols, phenols, lactones, naphthoic acid and phthalic acid that could be excreted into water (Keum et al 2006, Li et al 2006, Luan et al 2006, Seo et al 2006, Seo et al 2007, Wang and Zhao 2007). The metabolism of phenanthrene and pyrene in diatoms was carried mainly by the enzyme-oxidation

process converting PAHs to less or non-toxic forms of compounds (Wang and Zhao 2007). The application of diatoms in biosynthesis, biodegradation and biomineralization was gathered in Table 1.

#### **Diatoms and metal toxicity**

Soil and water contamination with toxic metal have become increasingly widespread and result in posing a global threat, which need more attention by scientists. Toxic metal pollution is a focus point of serious concern and the examination and monitoring water quality are becoming essential procedures (Ferreira da Silva et al 2009, Magnusson et al, Rudolph et al 2009, Wang and Wang 2008). Subcellular partitioning of metals may provide a mechanistic approach to investigate metal toxicity and tolerance. The presence of metal-tolerant populations at metal-contaminated sites has been shown for a number of genera of freshwater algae, including blue-green algae, filamentous greens and diatoms (Ivorra et al 2002). Diatoms are important bioindicators to monitor the metal concentrations in diverse habitats (Ferreira da Silva et al 2009, Magnusson et al Rudolph et al 2009, Wang and Wang 2008). If the purpose of toxicity tests is to predict the quantitative effect of a metal under particular field conditions, then the tests need to be refined. The assay medium should be similar to water at the field site with respect to factors such as chelating agents which influence toxicity (Whitton and Kelly 1995). The approach was applied by some researches to indicate sensitivity of microalgae for some ions shows the microalgae present in young biofilms were more sensitive to Zn or Cd than microalgae in old biofilms, the difference being apparent even when both types of biofilm were taken from a clean stream (Ivorra et al 2000). To examine seasons effects on the toxicity tolerance of microalgae to toxicants, in the River Ter, N-E. Spain microalgal communities at two different seasons (spring and summer) were tested for copper toxicity. Results of this study showed the tolerance to copper in microalgal communities was lower in spring than summer (Navarro et al 2002).

Algal species	Biosynthesis	Biodegradation	Biomineralization	Reference
Achnanthes oblongella Oestrup		waste		(Maznah and Mansor 2002)
Cocconeis placentula Ehr.		waste		(Maznah and Mansor 2002)
Fragilaria capucina Desm.		waste		(Maznah and Mansor 2002)
Chaetoceros cryptica	Carotenoids			(Bertrand 2010)
Haslea ostrearia	Marennine			(Robert <i>et al</i> 2002)
Melosira nummuloides			Platinum	(Yamazaki <i>et al</i> 2010)
Pinnularia sp.			Titanum	(Jeffryes <i>et al</i> 2008)
Pinnularia sp.			Cadmium	(Gutu <i>et al</i> 2009)

 Table 1. Diatom applications in biosynthesis, biodegradation and biomineralization

Human activities have been increasing the cadmium levels in soils and waters, disturbing many organisms in the primary trophic levels such as microalgae. Cadmium toxicity is due to Zn and Cu displacement in metalloenzymes and to the formation of reactive oxygen species (ROS), leading to oxidative stress (Goncalves et al 2009). Above normal concentration, ROS are potentially toxic and can react with lipids, proteins, photosynthetic pigments and nucleic acids (Moller et al 2007), leading to lipid peroxidation, decrease in chlorophyll and accessory pigment content (Sabatini et al 2009), membrane damage, inactivation of enzymes, DNA alterations and oxidation of proteins, thus affecting cell growth and viability (Qiu et al 2008). Cadmium (Cd) is a priority pollutant, and its toxicity is mainly related to binding to sulfhydryl groups of proteins or displacement of essential metals in metalloenzymes. Recently, a new approach, namely the subcellular partitioning model (SPM), which takes into account the cellular fates of metals, has been proposed to predict metal toxicity in aquatic organisms. The bioaccumulation, subcellular distribution, and toxicity of Cd in a marine diatom, Thalassiosira nordenskioeldii, under different irradiance levels was examined (Wang and Wang 2008). The findings showed that intracellular soluble Cd may be the best predictor of Cd toxicity under different nutrient conditions (Wang and Wang 2008). Besides, results of other studies showed that most Cd was distributed in the insoluble fraction (a combination of metal-rich granules, cellular debris, and organelles) in the diatom Thalassiosira weissflogii (Miao and Wang 2006). A good toxicity predictor should be rather constant under different environmental/ physiological conditions. Although good correlation between [Cd<sup>2+</sup>] and growth inhibition was observed among the different irradiance treatments, suggesting that differences in Cd toxicity cannot be entirely explained by [Cd<sup>2+</sup>], primarily because the diatoms displayed different bioaccumulation potentials (Desai et al 2006, Horvatic and Persic 2007, Miao and Wang 2006, Wang and Wang 2008).

Copper toxicity has recently been studied by many of researchers (Debelius et al 2009a, Debelius et al 2009b, Wang and Zheng 2008). Copper belongs to the category of "essential metals" and participates in growth, metabolism and enzyme activities. However, at high concentrations is toxic and have a direct and adverse influence on various physiological and biochemical processes (Debelius et al 2009a, Debelius et al 2009b). One of the most studied toxic effects of metals on microorganisms is growth inhibition. Attempts to standardize growth inhibition tests with microalgae for regulatory purposes have revealed a number of methodological problems. Most researchers suggested that the metal concentration that affects growth in microalgae, is largely variable and depends on the species used, cell density, composition of medium or physical culture

conditions (Franklin *et al* 2002a, Franklin *et al* 2002b, Wang and Zheng 2008).

Recently, copper toxicity in five marine microalgae, *Tetraselmis chuii, Rhodomonas salina, Chaetoceros sp., Isochrysis galbana and Nannochloropsis gaditana* were studied because they represent different classes of marine microalgae and are all easy to culture (Debelius *et al* 2009a). The results demonstrated that the degree of growth inhibition was affected in different ways, depending on the strain of microalgae and metal concentration added in solution. All of the strains studied showed an increase in the signals of side-angle light scatter, corresponding to an increase in cell size or change in shape, also could reflect cell complexity when exposed to the highest copper and lead concentrations of their study (Debelius *et al* 2009a, Debelius *et al* 2009b, Wang and Zheng 2008).

Significant relationships were detected between a number of measures of the diatom community and various heavy metals in the environment. The comparison between the reference and affected sites showed a shift from a diatom community dominated by *Nitzschia palea, Achnanthes minutissima* and *Amphora pediculus* to one dominated by *Nitzschia palea* and *Gomphonema parvulum* (Whitton and Kelly 1995).

All the results obtained from researches proved that toxicity of different metals and their effects on diatoms, cause to change in biological, physiological, and morphological changes in diatoms which can be used as a unique indicator for determination and investigation of metal toxicity.

## Conclusion

As it has been evidenced in the recent literature, diatoms are important synthesis microalgae for biomaterials. Additionally, they are appropriate indicators that can provide green tools in detecting contamination problems in aquatic environments because they are very sensitive to environmental stress. They also can be used in rehabilitation of suffered environment. However, until now, the aforementioned aspects remain unexplored for their quantitative analytical use and, as for example; it could be interesting to try the use of diatoms for trace element preconcentration media and as a cleaning step added after analyte determinations.

### **Ethical issues**

No ethical issues to be promulgated.

## **Conflict of interests**

No conflict of interests to be declared.

#### Acknowledgements

The financial support of the Research Center for Pharmaceutical Nanotechnology, Tabriz University of Medical Sciences, and Research Institute of Physiology and Biotechnology (RIPB) of Zanjan University are gratefully acknowledged.

#### References

Ashok P. **2009**. *Biotechnology for Agro-Industrial Residues Utilization*. Springer. 462 p.

Atazadeh I, Kelly MG, Sharifi M and Beardall J. **2009**. The effects of copper and zinc on biomass and taxonomic composition of algal periphyton communities from the River Gharasou, western Iran. *Oceanol Hydrobiol St*, 38(3), 3-14.

Atazadeh I and Sharifi M. **2010**. *Algae as Bioindicators*. Lambert Academic Publishing House.

Atazadeh I, Sharifi M and Kelly MG. **2007**. Evaluation of the Trophic Diatom Index for assessing water quality in River Gharasou, western Iran. *Hydrobiologia*, 589, 165-173.

Berard A, Dorigo U, Humbert JF, Leboulanger C and Seguin F. **2002**. Application of the Pollution-Induced Community Tolerance (PICT) method to algal communities: its values as a diagnostic tool for ecotoxicological risk assessment in the aquatic environment. *Ann Limnol*, 38(3), 247-261.

Bertrand M. **2010**. Carotenoid biosynthesis in diatoms. *Photosynth Res*, 106(1-2), 89-102

Brunner E, Gröger C, Lutz K, Richthammer P, Spinde K and Sumper M. **2009**. Analytical studies of silica biomineralization: towards an understanding of silica processing by diatoms. *Appl Microbiol Biotechnol*, 84(4), 607-16.

Chen X, Ostadi H and Jiang K. **2010**. Three-dimensional surface reconstruction of diatomaceous frustules. *Anal Biochem*, 403, 63-66.

Debelius B, Forja JM, DelValls A and Lubian LM. **2009a**. Toxicity and bioaccumulation of copper and lead in five marine microalgae. *Ecotoxicol Environ Saf*, 72(5), 1503-1513.

Debelius B, Forja JM, DelValls TA and Lubian LM. **2009b**. Toxicity of copper in natural marine picoplankton populations. *Ecotoxicology*, 18(8), 1095-1103.

Desai SR, Verlecar XN, Nagarajappa and Goswami U. **2006**. Genotoxicity of cadmium in marine diatom Chaetoceros tenuissimus using the alkaline Comet assay. *Ecotoxicology*, 15(4), 359-363.

Dolatabadi JEN and de la Guardia M. **2011**. Applications of diatoms and silica nanotechnology in biosensing, drug and gene delivery, and formation of complex metal nanostructures. *Trac-Trends in Analytical Chemistry*, 30(9), 1538-1548.

Evans GM and Furlong JC. **2003**. *Environmental Biotechnology, Theory and Application*. West Sussex: John Wiley & Sons.

Ferreira da Silva E, Almeida SF, Nunes ML, Luis AT, Borg F, Hedlund M *et al.* **2009**. Heavy metal pollution downstream the abandoned Coval da Mo mine (Portugal) and associated effects on epilithic diatom communities. *Sci Total Environ*, 407(21), 5620-5636.

Florence TM and Stauber JL. **1986**. Toxicity of Copper Complexes to the Marine Diatom Nitzschia Closterium. *Aquat Toxicol*, 8, 11-26.

Franklin NM, Stauber JL, Apte SC and Lim RP. **2002a**. Effect of initial cell density on the bioavailability and toxicity of copper in microalgal bioassays. *Environ Toxicol Chem*, 21(4), 742-751.

Franklin NM, Stauber JL, Lim RP and Petocz P. **2002b**. Toxicity of metal mixtures to a tropical freshwater alga (Chlorella sp): the effect of interactions between copper, cadmium, and zinc on metal cell binding and uptake. *Environ Toxicol Chem*, 21(11), 2412-2422.

Glazer AN and Hiroshi N. **2007**. *Microbial Biotechnology*. Cambridge: Cambridge University Press. 577 p.

Goncalves JF, Tabaldi LA, Cargnelutti D, Pereira LB, Maldaner J, Becker AG *et al.* **2009**. Cadmium-induced oxidative stress in two potato cultivars. *Biometals*, 22(5), 779-792.

Gordon R, Sterrenburg FAS and Sandhage KH. **2005**. A Special Issue on Diatom Nanotechnology. *J Nanosci Nanotech*, 5, 4.

Gutu T, Debra KG, Jeffryes C, Gregory LR and Jiao J. **2009**. Electron microscopy and optical characterization of cadmium sulphide nanocrystals deposited on the patterned surface of diatom biosilica. *J Nanomater*, doi:10.1155/2009/860536.

Haynes K, Hofmann TA, Smith CJ, Ball AS, Underwood GJ and Osborn AM. **2007**. Diatom-derived carbohydrates as factors affecting bacterial community composition in estuarine sediments. *Appl Environ Microbiol*, 73(19), 6112-6124.

Heredia A, van der Strate HJ, Delgadillo I, Basiuk VA and Vrieling EG. **2008**. Analysis of organo-silica interactions during valve formation in synchronously growing cells of the diatom Navicula pelliculosa. *Chembiochem*, 9(4), 573-584.

Hess P. **2010**. Requirements for screening and confirmatory methods for the detection and quantification of marine biotoxins in end-product and official control. *Anal Bioanal Chem*, 397(5), 1683-1694.

Hildebrand M. **2008**. Diatoms, Biomineralization Processes, and Genomics. *Chem Rev*, 108(11),4855-74.

Horvatic J and Persic V. **2007**. The effect of  $Ni^{2+}$ ,  $Co^{2+}$ ,  $Zn^{2+}$ ,  $Cd^{2+}$  and  $Hg^{2+}$  on the growth rate of marine diatom Phaeodactylum tricornutum Bohlin: microplate growth inhibition test. *Bull Environ Contam Toxicol*, 79(5), 494-498.

Ivorra N, Barranguet C, Jonker M, Kraak MHS and Admiraal W. **2002**. Metal-induced tolerance in the freshwater microbenthic diatom Gomphonema parvulum. *Envir Pollut*, 116, 147-157.

Ivorra N, Bremer S, Guasch H, Kraak MHS and Admiraal W. **2000**. Differences in the sensitivity of benthic microalgae to Zn and Cd regarding biofilm development and exposure history. *Environ Toxicol Chem*, 19, 1332-1339.

Jeffryes C, Gutu T, Jiao J and Rorrer GL. **2008**. Metabolic insertion of nanostructured TiO2 into the patterned biosilica of the diatom Pinnularia sp. by a two-stage bioreactor cultivation process. *ACS Nano*, 2(10), 2103-2112.

Kelly MG. **1998**. Use of the trophic diatom index to monitor eutrophication in rivers. *Water Res*, 32, 236-242.

Keum YS, Seo JS, Hu Y and Li QX. **2006**. Degradation pathways of phenanthrene by Sinorhizobium sp. C4. *Appl Microbiol Biotechnol*, 71(6), 935-941.

Li H, Liu YH, Luo N, Zhang XY, Luan TG, Hu JM *et al.* **2006**. Biodegradation of benzene and its derivatives by a psychrotolerant and moderately haloalkaliphilic Planococcus sp. strain ZD22. *Res Microbiol*, 157(7), 629-636.

Liu Y, Guan Y, Gao Q, Tam NF and Zhu W. **2010**. Cellular responses, biodegradation and bioaccumulation of endocrine disrupting chemicals in marine diatom Navicula incerta. *Chemosphere*, 80(5), 592-599.

Losic D, Mitchell JG and Voelcker NH. **2009**. Diatomaceous Lessons in Nanotechnology and Advanced Materials. *Adv Mater*, 21, 2947-2958.

Luan TG, Yu KS, Zhong Y, Zhou HW, Lan CY and Tam NF. **2006**. Study of metabolites from the degradation of polycyclic aromatic hydrocarbons (PAHs) by bacterial consortium enriched from mangrove sediments. *Chemosphere*, 65(11), 2289-2296.

Magnusson M, Heimann K, Quayle P and Negri AP. **2010**. Additive toxicity of herbicide mixtures and comparative sensitivity of tropical benthic microalgae. *Mar Pollut Bull*, 60 (11), 1978–1987.

Marchetti A and Cassar N. **2009**. Diatom elemental and morphological changes in response to iron limitation: a brief review with potential paleoceanographic applications. *Geobiology*, 7(4), 419-431.

Maznah WOW and Mansor M. **2002.** Aquatic pollution assessment based on attached diatom communities in the Pinang River Basin, Malaysia. *Hydrobiologia*, 487(1), 229-241.

Miao AJ and Wang WX. **2006**. Cadmium toxicity to two marine phytoplankton under different nutrient conditions. *Aquat Toxicol*, 78(2), 114-126.

Mohan BS and Hosetti BB. **1999**. Aquatic Plant for Toxicity Assessment. *Environ Res A*, 81, 259-274.

Moller IM, Jensen PE and Hansson A. **2007**. Oxidative modifications to cellular components in plants. *Annu Rev Plant Biol*, 58, 459-481.

Navarro E, Baun A, Behra R, Hartmann NB, Filser J, Miao AJ *et al.* **2008**. Environmental behavior and ecotoxicity of engineered nanoparticles to algae, plants, and fungi. *Ecotoxicology*, 17(5), 372-386.

Navarro E, Guasch H and Sabater S. **2002**. Use of microbenthic algal communities in ecotoxicological tests for the assessment of water quality: the Ter river case study. *J Appl Phycol*, 13, 114-120.

Pepper IL and Gerba CP. **2005**. *Environmental Microbiology*. Edition S, editor. San Diego, California: Elsevier. 226 p.

Qiu RL, Zhao X, Tang YT, Yu FM and Hu PJ. **2008**. Antioxidative response to Cd in a newly discovered cadmium hyperaccumulator, Arabis paniculata F. *Chemosphere*, 74(1), 6-12. Robert J-M, Morançais M, Pradier E, Mouget J-L and Tremblin2 G. **2002**. Extraction and quantitative analysis of the blue-green pigment "marennine" synthesized by the diatom Haslea ostrearia. *J Appl Phyco*, 14(4), 299-305.

Rudolph A, Medina P, Urrutia C and Ahumada R. **2009**. Ecotoxicological sediment evaluations in marine aquaculture areas of Chile. *Environ Monit Assess*, 155(1-4), 419-429.

Saade A and Bowler C. **2009**. Molecular Tools for Discovering the Secrets of Diatoms. *BioScience*, 59(9), 757-765.

Sabatini SE, Juarez AB, Eppis MR, Bianchi L, Luquet CM and Rios de Molina Mdel C. **2009**. Oxidative stress and antioxidant defenses in two green microalgae exposed to copper. *Ecotoxicol Environ Saf*, 72(4), 1200-1206.

Schindler DW. **1987**. Detecting ecosystem responses to anthropogenic stress. *Can J Fish Aquat*, 44, 6-25.

Schroder HC, Brandt D, Schlossmacher U, Wang X, Tahir MN, Tremel W *et al.* **2007**. Enzymatic production of biosilica glass using enzymes from sponges: basic aspects and application in nanobiotechnology (material sciences and medicine). *Naturwissenschaften*, 94(5), 339-359.

Seo JS, Keum YS, Hu Y, Lee SE and Li QX. **2006**. Phenanthrene degradation in Arthrobacter sp. P1-1: initial 1,2-, 3,4- and 9,10-dioxygenation, and meta- and ortho-cleavages of naphthalene-1,2-diol after its formation from naphthalene-1,2-dicarboxylic acid and hydroxyl naphthoic acids. *Chemosphere*, 65(11), 2388-2394.

Seo JS, Keum YS, Hu Y, Lee SE and Li QX. **2007**. Degradation of phenanthrene by Burkholderia sp. C3: initial 1,2- and 3,4-dioxygenation and meta- and ortho-cleavage of naphthalene-1,2-diol. *Biodegradation*, 18(1), 123-131.

Srinivas T. **2008**. *Environmental Biotechnology*. New Delhi: New Age International.

Wang L and Zheng B. **2008**. Toxic effects of fluoranthene and copper on marine diatom Phaeodactylum tricornutum. *J Environ Sci (China)*, 20(11), 1363-1372.

Wang M and Wang WX. **2008**. Cadmium toxicity in a marine diatom as predicted by the cellular metal sensitive fraction. *Environ Sci Technol*, 42(3), 940-946.

Wang XC and Zhao HM. **2007**. Uptake and Biodegradation of Polycyclic Aromatic Hydrocarbons by Marine seaweed. *J coastal res*, SI(50), 56–1061.

Whitton BA and Kelly MG. **1995**. Use of algae and other plants for monitoring rivers. *Aust J Ecol*, 20, 45-56.

Yamazaki T, Sasanuma H, Mayama S and Umemura K. **2010**. Cultivation of Melosira nummuloides cells in the presence of platinum: Preparation of metal-containing frustules. *Phys Status Solidi*, 7(11-12), 2759-2762.

Yuan P, Yang D, Lin Z, He H, Wen X, Wang L *et al.* **2006**. *J Non-Cryst Solids*, 352, 3762-3771.