

Antioxidant, Total Phenol and Flavonoid Contents of Two *Pedicularis* L. Species from Eastern Azerbaijan, Iran

Laleh Khodaie^{1,2}, Sedigheh Bamdad¹, Abbas Delazar¹, Hossein Nazemiyeh^{3*}

¹Drug Applied Research Center, Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran ²Students' Research Committee, Tabriz University of Medical Sciences, Tabriz, Iran ³Research Center for Pharmaceutical Nanotechnology, Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran

ARTICLE INFO

ABSTRACT

Article Type: Research Article

Article History: Received: 13 Feb 2012 Revised: 6 March 2012 Accepted: 20 March 2012 ePublished: 24 March 2012

Keywords: Pedicularis sibthorpii Pedicularis wilhelmsiana Antioxidants Polyphenols Flavonoids Introduction: Pedicularis sibthorpii and P. wilhelmsiana are endemic species mainly found in North-West of Iran. Plants of genus Pedicularis produce some important polyphenols and flavonoids. In the present work, total phenol and flavonoid contents of the mentioned species as well as their antioxidant capacity have been evaluated. Methods: Methanol extract of samples was fractionated by SPE method using an ODS cartridge and their ¹H-NMR spectra were recorded. Total phenols and flavonoids of methanol extracts were determined using Folin- Ciocalteu and aluminum chloride methods. For determining antioxidant activity of the extracts and fractions, bleaching of purple color methanol solution of 1, 1-diphenylpycryl hydrazyl (DPPH) was measured by spectrophotometric assay. Results: Total phenols of Pedicularis sibthorpii and P. wilhelmsiana were in the range of 8-30 mg g⁻¹ and 9-20 mg g⁻¹, respectively. The 40% and 60% fractions of *P. sibthorpii* and the 20%, 40% and 60% fractions of P. wilhelmsiana showed higher amounts of phenolic compounds. The total flavonoid contents of P. sibthorpii and P. wilhelmsiana were in the range of 0-215 mg g⁻¹ and 0-177 mg g⁻¹, respectively, whereas the 40% and 60% fractions showed higher flavonoid amounts. Antioxidant activity of P. sibthorpii and P. wilhelmsia*na* were in the range of 0.01-0.7 mg mL⁻¹ and 0.01-1.02 mg mL⁻¹. In the same manner, the 20% and 40% fractions of P. sibthorpii and the 40% and 60% fractions of P. wilhelmsiana had lower RC₅₀ than that of other fractions. Conclusion: Fractions with lower RC₅₀ had higher contents of phenolic and flavonoid compounds. The results of NMR spectra were parallel with these findings and show that it is worth to do phytochemical studies on P. sibthorpii and P. wilhelmsiana.

Introduction

Free radicals can affect whole body system through contribution to many kinds of degenerative diseases including diabetes and cardiovascular damage (Yang *et al* 2010), CNS injury (Cadet 1998, Demopoulos *et al* 1982, Demopoulos *et al* 1980), cancer (Samra *et al* 2011, Vera-Ramirez *et al* 2011), liver and kidney damage (Muriel 2009, Small *et al* 2012), atherosclerosis, inflammatory joint disease, asthma (Rashidi *et al* 2010), gastritis (Salim 1992) and so on. Oxidation process is one of the most important routs for producing free radicals in food, drugs and even living systems (Mc Cord 2000).

Discovery of free radicals' impacts on biological system has led to the free-radical theory of aging which implies that preventing free radicals' action could influence lifespan. Though the healthy body produces a range of its own antioxidant to overwhelm free radicals, supplement natural antioxidants as free radical scavengers may be needed to boost the defensive system and slow down aging. This postulation has evoked great efforts for finding the powerful free radical scavengers to overcome harmful effects of oxidative stress. Recently attentions have been focused on the therapeutic potential of green foods and medicinal plants which is believed to reduce free radical induced tissue injury by trapping them (Beta *et al* 2005, Cai *et al* 2004, Chen *et al* 2007, Dudonne *et al* 2009). Higher plants produce a variety of antioxidant compounds of which, polyphenols are assumed to be the most potent one (Wang *et al* 2007, Guangrog *et al* 2008).

The genus *Pedicularis* produces iridoids, phenylethanoids, phenylpropanoids and flavonoids (Akdemir *et al* 1991, Fujii *et al* 1995, Liu *et al* 1991, Su *et al* 1998) which their antioxidant properties have been established, previously (Scalbert *et al* 2005). Majority of the *Pedicu*-

*Corresponding author: Hossein Nazemiyeh (PhD), Tel.: +98-411-3367914, Fax: +98-411-3367929, E-mail: nazemiyehh@yahoo.com Copyright © 2012 by Tabriz University of Medical Sciences laris species are widely distributed in China (Hanbi et al 1998) and most phytochemical studies have been carried out on these species. Pedicularis species have different therapeutic applications in Traditional Chinese Medicine (Jiang et al 2003, Zhang et al 2008). As part of our ongoing research on bioactive constituents of Iranian medicinal plants, we focused on native Pedicularis species. Pedicularis genus is represented by 9 species in flora of Iran (Wendelbo 1981) which their medicinal uses are unknown. Pedicularis sibthorpii and P. wilhelmsiana grow in Azerbaijan province, Iran and according to the best of our knowledge, their biological effects and chemical constituents are not investigated, yet. In the present work, the results of DPPH assay, total phenol and flavonoid content of P. sibthorpii Boiss and P. wilhelmsiana Fisch. extracts and their relationship with ¹H-NMR spectra have been reported.

Materials and methods

Plant materials

The samples of *Pedicularis sibthorpii* Boiss. were collected from Lighvan valley -Tabriz and samples of *P. wilhelmsiana* Fisch. ex M. Bieb. were collected from Arasbaran region in East Azerbaijan province, Iran in 2009. Voucher specimens for these collections (*P. wilhelmsiana*, TUM- FPh 700; *P. sibthorpii*, TUM- FPh 701) have been deposited in the Herbarium of the Faculty of Pharmacy, Tabriz - Iran.

Extraction and fractionation

Powdered air-dried aerial parts of plants (200g) were extracted by n-hexane (8 h), dichloromethane (10 h) and methanol (8 h) using a Soxhlet, respectively. Solvents were removed *in vacuo* by rotary evaporator at an ambient temp. Further studies were carried out on methanol extract fractions prepared by solid phase extraction (SPE) method.

The methanol extract (2 g) was loaded on a Sep-pak (Vac 35 cc; 10g; C18 cartridge) cartridge and fractions were eluted by step gradients of methanol- water mixtures (fr1,10% methanol; fr2, 20% methanol; fr3, 40% methanol; fr4, 60% methanol; fr5, 80% methanol; fr6, 100% methanol; 200 mL each). The solvents of eluted fractions were removed in vacuo and 40°C.

DPPH assay

Antioxidant activity of the extracts and fractions was measured according to Asnaashari *et al* (2011) with some modifications. In order to obtain dilutions, different sample concentrations were prepared in methanol and 5 mL of each concentration were added to 5mL of 0.004% methanolic solution of DPPH. After completion of reaction at room temperature for 30 min, bleaching of DPPH was monitored at 517 nm against a blank. Inhibition of DPPH was calculated as RC_{50} , extrapolated from dose-response curve. Tests were carried out in triplicate and the same procedure was applied for the quercetin as a positive control.

Determination of total phenol content

Determination of total phenol content was carried out by Folin- Ciocalteu test according to Chun *et al* (2003) with some modifications. One mL of the extract and fractions' sample (solved in 60% acetone, 5 mg/100 mL) was mixed with 200 μ l Folin- Ciocalteu reagent and 1 mL of aqueous Na₂CO₃. The mixtures were left at room temperature for 30 min and the phenol contents were determined by colorimetric method at 715 nm. The calibration curve was prepared using Gallic acid solutions at concentrations of 1- 0.01562 mg/mL in 60% acetone. Total phenol contents were expressed in terms of gallic acid equivalent (mg g⁻¹).

Determination of total flavonoids content

The total flavonoid content was estimated using aluminum chloride colorimetric assay (Zhishen *et al* 1999, Zou *et al* 2004). The 0.5 mL of test samples' solution in methanol (5mg/100mL) were mixed with 2mL of distilled water and 150 μ l of 5% sodium nitrate. After 6 min, 150 μ l of 10% aluminum chloride and 2mL of 1 M sodium hydroxide were added and left at room temperature for 15 min. Absorbance of the mixtures was measured at 510 nm (UV-Visible Ultraspec 2000 spectrophotometer, England) and total flavonoid contents were calculated as rutoside equivalents from a calibration curve of rutoside. The calibration curve was prepared in the same manner using 0.01562-1 mg/mL of rutoside solutions in methanol.

NMR spectra from methanolic fractions

NMR spectra were recorded in CD₃OD on a Bruker 200 MHz NMR spectrometer. TMS was used as the internal standard.

Results and discussion

According to the literature records, chemical constituents and bioactivities of different *Pedicularis* species have previously been investigated. Iridoids, phenylethanoids, phenylpropanoids and flavonoids are the major secondary metabolites of the genus *Pedicularis* (Akdemir *et al* 1991, Fujii *et al* 1995, Liu *et al* 1991, Su *et al* 1998). The antioxidant effects of phenylpropanoid glycosides, widely distributed in genus *Pedicularis*, were studied in different models (Zheng *et al* 1993, Scalbert *et al* 2005, Hosoya *et al* 2008, Ahmad *et al* 2009, Biao *et al* 2009, Zhu *et al* 2010).

Polyphenols consisting of a wide range of biogenic molecules play numerous roles in living organisms. Flavonoids, as the main class of polyphenols, widely distributed in plants, display numerous pharmacological effects (Rashidi *et al* 2010). It has been shown that free radical scavenging property of flavonoids is responsible for these effects (Cotelle *et al* 1996, Brown *et al* 1998, Seyoum *et al* 2006, Tsimogiannis *et al* 2006). A common and rapid method to evaluate free radical scavenging ability of flavonoids involves the use of stable free radical 2,2-diphenyl-1-picrylhydrazyl (DPPH).

Extracts of *P. sibthorpii* and *P. wilhelmsiana* and their fractions exhibited varying degrees of free radical scavenging capacities as determined by DPPH methods (Table 1 and Table 2). Total extract of *P. sibthorpii* (RC_{50} = 0.033 mg mL⁻¹) showed stronger radical scavenging effect than that of *P. wilhelmsiana* (RC_{50} = 0.15 mg mL⁻¹). Considering the RC_{50} of quercetin (2.92 x 10⁻⁵ mg mL⁻¹), it could be a moderate effect. The experiments showed that the 20% and 40% Sep-Pak fractions of *P. sibthorpii* as well as the 40% and 60% Sep-Pak fractions of *P. wilhelmsiana* have lower RC_{50} than that of other fractions.

Total phenols were assessed by Folin Ciocalteu method and results were reported as gallic acid equivalents by reference to standard curve (y = 0.0282+0.0043, $r^2=0.998$). Total phenols of *P. sibthorpii* and *P. wilhelmsiana* and their fraction were demonstrated in Table 1 and Table 2. According to these findings, 40% and 60% Sep-Pak fractions revealed the highest amount of total phenols at 30.06 and 25.07 mg g⁻¹ respectively, approximately 2 times more than methanol extracts.

Flavonoid content was determined by aluminum chloride method and expressed as rutoside equivalents in mg g⁻¹ dry extracts and fractions using a standard curve of rutoside (y= 2.0903x- 0.0298, r²= 0.998). As can be seen from the Table 1 and Table 2, flavonoid content of 40% and 60% Sep-Pak fractions was superior to the flavonoid content of other fractions and extracts. The lowest flavonoid content was found within 20%, 80% and 100% fractions.

The collected data have shown the existence of a good harmony between antioxidant potency of samples and their total phenolic and flavonoid content. The maximum antioxidant, total polyphenols and flavonoid content were seen in 40% and 60% Sep-Pak fractions whereas the other fractions were found to contain nearly low or neglectable amounts. Generally, the higher antioxidant activity of the mentioned fractions might be attributed to their contents of total polyphenols and especially flavonoids. However according to Prior *et al* (2005), many other compounds may contribute to the reaction and cause a false positive error.

In the present study, the issue was partly clarified by using ¹H- NMR spectra as fingerprints to indicate significant differences between the chemical constituents of fractions (Figures 1 and 2). The ¹H- NMR spectra of

10% Sep-Pak fractions belonging to P. sibthorpii and P. wilhelmsiana (Figures 1-a and 2-a, respectively) have revealed that there is no phenolic compound, but iridoids (signals in the δ 3-6 ppm) in this fraction. In the same way, the Figures 1-b and 2-b show that some phenolic compounds such as phenylethanoids and flavonoids may exist in low concentrations in 20% fractions of P. sibthorpii and P. wilhelmsiana. Verbascoside is a common phenylpropanoid of the *Pedicularis* genus and its signals have clearly been seen in the ¹H-NMR spectra of 40% fraction (Figures 1-c and 2-c). Flavonoids may exist in 40% fractions in lower concentration, but their signals were covered by verbascoside signals due to its high concentration. Figures 1-d and 2-d, which are NMR spectra of 60% fraction, indicate that phenolic compounds such as phenylethanoids and flavonoids exist in this fraction. Obviously phenolic compounds exist in very low concentrations in 80% fractions (Figures 1-e and 2-e) and there is no sign of similar compounds in 100% fractions (Figures 1-f and 2-f).

According to these finding, it can be concluded that there is a relationship between antioxidant activity, phenolic and flavonoid content in these two species, because fractions with lower RC₅₀ have higher contents of phenolic and flavonoid compounds. The results of NMR spectroscopy are parallel with these findings, too. Fractions which show signals in aromatic regions (δ 6-8 ppm) possess higher polyphenolic compounds and lower RC50.

 Table1. Antioxidant, total phenol and flavonoid contents of P. sibthorpii

Extract or	Total phenol content	Flavonoid	Antioxidant activity ^a
fractions	(as gallic acid equiva-	content	(RC ₅₀ ; mg mL ^{−1})
	lents) mg g⁻¹	mg g ⁻¹	
Total extract	15.28	47.50	0.0330
10%	08.56	01.80	00.820
20%	12.59	00.00	00.070
40%	30.06	118.61	00.012
60%	25.07	214.76	00.107
80%	10.74	00.00	00.337
100%	10.01	00.00	00.616
0		E	

^a Positive control (quercetin): 2.92 x 10⁻⁵

 Table 2. Antioxidant, total phenol and flavonoid contents of P.

 wilhelmsiana

Extract or	Total phenol content	Flavonoid	Antioxidant activity ^a
fractions	(as gallic acid equi-	content mg g ⁻¹	$(RC_{50}; mg mL^{-1})$
	valents) mg g ⁻¹		
Total extract	12.18	35.60	00.15
10%	09.17	01.55	03.82
20%	17.52	00.00	00.18
40%	19.31	154.14	0.017
60%	18.43	176.60	00.0740
80%	11.32	00.00	00.1774
100%	09.84	00.00	01.0192

^a Positive control(quercetin): 2.92 x 10⁻⁵



Fig. 1. ¹H-NMR spectra of Sep-Pak fraction obtained from *P. sibthropii* methanolic extract. The spectra were recorded in CD3OD. a) 10% fraction, b) 20% fraction, c) 40% fraction, d) 60% fraction, e) 80% fraction and f) 100% fraction.



Fig. 2. ¹H-NMR spectra of Sep-Pak fraction obtained from *P. wilhelmsiana* methanolic extract. The spectra were recorded in CD₃OD. a) 10% fraction, b) 20% fraction, c) 40% fraction, d) 60% fraction, e) 80% fraction and f) 100% fraction.

Conclusion

Increasing demands for natural antioxidant have provoked great efforts for finding new and potent free radical scavengers from plants. *Pedicularis* species yield a series of antioxidant, antitumor, antifungal, antidiabetic and anti-inflammatory secondary metabolites (Beta *et al* 2005, Cai *et al* 2004). According to some evidences, phenylpropanoids and flavonoids are responsible for antioxidant effects in plants (Choudhary *et al* 2011).

The results of present work show that it is worth to do phytochemical studies on Iranian *Pedicularis* species and purify their antioxidant compounds which are estimated to be phenylethanoids and flavonoids according to the ¹H-NMR findings. It could be concluded that fractionation of extracts and running their H-NMR could be a valuable method for predicting and comparing their anti-oxidant capacity.

Ethical issues

Not applicable in this research.

Conflict of interests

Authors declared no conflicts of interests.

References

Ahmad I, Ahmad N and Wang F. **2009**. Antioxidant phenylpropanoid glycosides from *Buddleja davidii*. *Journal of Enzyme Inhibition and Medicinal Chemistry*, 24(4), 993-997.

Akdemir Z, Çali II and Junior P. **1991**. Iridoids and phenylpropanoid glycosides from *Pedicularis nordmanniana*. *Planta Medica*, 57(6), 584-585.

Asnaashari S, Dadizadeh E, Talebpour AH, Eskandani M and Nazemiyeh H. **2011**. Free Radical Scavenging Potential and Essential Oil Composition of the *Dorema glabrum* Fisch. C.A. Mey Roots from Iran. *BioImpacts*, 1(4), 241-244.

Beta T, Names JED and Sapirstein HD. **2005**. Phenolic content and antioxidant activity pearled wheat and roller-milled fractions. *Cereal Chemistry*, 82(4), 390-393.

Biao CH, Hua TN and Sheng PC. **2009**. Progress in research on *Pedicularis* plants. *China Journal of Chinese Materia Medica*, 34(19), 2536-2546.

Brown JE, Khodr H, Hider RC and Rice-Evans CA. **1998**. Structural dependence of flavonoid interactions with Cu2+ ions: implications for their antioxidant properties. *Biochemical Journal*, 15 (330), 1173-1178.

Cadet JL. **1988**. Free radical mechanisms in the central nervous system: an overview. *International Journal of Neuroscience*, 40(1-2), 13-18.

Cai Y, Luo Q, Sun M and Corke H. **2004**. Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. *Life Science*, 74, 2157-2184.

Chen HY and Yen GC. **2007**. Antioxidant activity and free radical scavenging capacity of extracts from guava (*Psidium guajava* L.) leaves. *Food Chemistry*, 101, 686-694.

Choudhary RK and Swarnkar PL. **2011**. Antioxidant activity of phenolic and flavonoid compounds in some medicinal plants of India. *Natural Product Research*, 25(11), 1101-1109.

Chun OK, Kim DO and Lee CY. **2003**. Superoxide radical scavenging activity of the major polyphenols in fresh plums. *Journal of Agriculture and Food Chemistry*, 51, 8067-8072.

Cotelle N, Bernier JL, Catteau JP, Pommery J, Wallet JC and Gaydou EM. **1996**. Antioxidant properties of hydroxy-flavones. *Free Radical Biology and Medic*ine, 20(1), 35-43.

Demopoulos HB, Flamm ES, Pietronigro DD and Seligman ML. **1980**. The free radical pathology and the microcirculation in the major central nervous system disorders. *Acta Physiologica Scandinavica Supplement*, 492, 91-119.

Demopoulos HB, Flamm ES, Seligman ML, Pietronigro DD, Tomasula J and DeCrescito V. **1982**. Further studies on freeradical pathology in the major central nervous system disorders: effect of very high doses of methylprednisolone on the functional outcome, morphology, and chemistry of experimental spinal cord impact injury. *Canadian Journal of Physiology and Pharmacology*, 60(11), 1415-1424.

Dudonne S, Vitrac X, Coutiere P, Woillez M and Merillon JM. **2009**. Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial interest using DPPH, ABTS, FRAP, SOD, and ORAC assays. *Journal of Agriculture and Food Chemistry*, 57(5), 1768-1774.

Fujii M, Miyaichi Y and Tomimori T. **1995**. Flavonoid, phenylethanoid and iridoid constituents of the whole plant of *Pedicularis longiflora var. tubiformis. Planta Medica*, 61(6), 584.

Guangrog H, Jiaxin J and Dehui D. **2008**. Antioxidative and antibacterial activity of the methanol extract of *Artemisia anomala* S. Moore. *African Journal of Biotechnology*, 7, 1335-1338.

Hanbi Y, Holmgren NH and Mill RR. **1998**. *Pedicularis*. In: *Flora of China*, Wu CY and Raven PH. (eds.), Vol. 18, Science Press, Beijing, 97-209.

Hosoya T, Yun YS and Kunugi A. **2008**. Antioxidant phenylpropanoid glycosides from the leaves of *Wasabia japonica*. *Phytochemistry*, 69, 827-832

Jiang TF, Ou QY and Shi YP. **2003**. Separation and determination of phenylpropanoid glycosides from *Pedicularis* species by capillary electrophoresis. *Journal of Chromatography A*, 986(1), 163-167.

Lee OH and Lee BY. **2010**. Antioxidant and antimicrobial activities of individual and combined phenolics in *Olea europaea* leaf extract. *Bioresource Technology*, 101(10), 3751-3754.

Liu ZM and Jia ZJ. 1991. Phenylpropanoid and iridoid glycosides from *Pedicularis striata*. *Phytochemistry*, 30(4), 1341-1344.

McCord JM. **2000**. The evolution of free radicals and oxidative stress. *American Journal of Medicine*, 108(8), 652-659.

Muriel P. **2009**. Role of free radicals in liver diseases. *Hepatology International*, 3(4), 526-536.

Prior RL, Wu X and Schaich K. **2005**. Standardized Methods for the determination of antioxidant capacity and phenolics in foods and dietary supplements. *Journal of Agriculture and Food Chemistry*, 53, 4290-4302.

Rashidi MR and Nazemiyeh H. **2010**. Inhibitory effects of flavonoids on molybdenum hydroxylases activity; a review. *Expert Opinion on Drug Metabolism & Toxicolog*, 6(2), 133-152.

Salim AS. **1992**. Oxygen-derived free radical scavengers protect patients against the complications of erosive gastritis. *Intensive Care Medicine*, 18(1), 61-62.

Samra ZQ, Pervaiz S, Shaheen S, Dar N and Athar MA. **2011**. Determination of oxygen derived free radicals producer (xanthine oxidase) and scavenger (paraoxonase1) enzymes and lipid parameters in different cancer patients. *Clinical Laboratory*, 57(9-10), 741-747.

Scalbert A., Johnson IT and Saltmarsh M. 2005. Polyphenols: antioxidants and beyond. *American Journal of Clinical Nutrition*, 81(suppl), 215S-217S.

Seyoum A., Asres K and El-Fiky FK. **2006**. Structure–radical scavenging activity relationships of flavonoids. *Phytochemistry*, 67, 2058-2070.

Small DM, Coombes JS, Bennett N, Johnson DW and Gobe GC. **2012.** Oxidative stress, anti-oxidant therapies and chronic kidney disease. *Nephrology* (*Carlton*), Jan 31, doi: 10.1111/j.1440-1797.2012.01572.x.

Su BN, Ma LP and Jia ZJ. **1998**. Iridoid and phenylpropanoid glycosides from *Pedicularis artselaeri*. *Planta Medica*, 64(8), 720-723.

Tsimogiannis DI and Oreopoulou V. **2006**. The contribution of flavonoid C-ring on the DPPH free radical scavenging efficiency. A kinetic approach for the 3',4'-hydroxy substituted members. *Innovative Food Science and Emerging Technologies*, 7, 140-146.

Valko M, Rhodes CJ, Moncol J, Izakovic M and Mazur M. **2006**. Free radicals, metals and antioxidants in oxidative stressinduced cancer. *Chemico- Biological Interactions*, 160(1), 1-40.

Vera-Ramirez L, Sanchez-Rovira P, Ramirez-Tortosa MC, Ramirez-Tortosa CL, Granados-Principal S, Lorente JA *et al.* **2011**. Free radicals in breast carcinogenesis, breast cancer progression and cancer stem cells. Biological bases to develop oxidative-based therapies. *Critical Reviews Oncology / Hematology*, 80(3), 347-368.

Wang Y, Yin J, Qiao Y, Zhang H and Lu X. **2007**. Studies on antioxidant activity and chemical constituents of *Artemisia* halodendron. Asian Journal of Traditional Medicines, 2, 30-33.

Wendelbo P. **1981**. *Pedicularis* In: *Flora Iranica*, Rechinger KH (editor), No. 147, Akademische Druk- u. Verlagsanstalt, Graz, 189-207.

Yang Y, Hayden MR, Sowers S, Bagree SV and Sowers JR. **2010**. Retinal redox stress and remodeling in cardiometabolic syndrome and diabetes. *Oxidative Medicine and Cellular Longevity*, 3(6), 392-403.

Zhang L, Ravipati AS, Koyyalamudi SR, Jeong SC, Reddy N, Smith PT *et al.* **2011**. Antioxidant and Anti-inflammatory Activities of Selected Medicinal Plants Containing Phenolic and Flavonoid Compounds. *Journal of Agriculture and Food Chemistry*, 59(23), 12361-12367.

Zhang ZX, xie WD and Jia ZJ. **2008**. Glycosids from two *Pedicularis* species. *Biochemical Systematics and Ecology*, 36(5-6), 467-472.

Zheng RL, Wang PF, Li J, Liu ZM and Jia ZJ. **1993**. Inhibition of the autoxidation of linoleic acid by phenylpropanoid glycosides from *Pedicularis* in micelles. *Chemistry and Physics of Lipids*, 65(2), 151-154.

Zhishen J, Mengcheng T and Jianming W. **1999**. The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. *Food Chemistry*, 64, 555-559.

Zhu M, Tan N, Zhu H, Zeng G, He W, Yu B, Chen X. **2010**. Anti-sports anaemia effects of verbascoside and martynoside in mice. *International Journal of Sports Medicine*, 31(8), 537-541.

Zou Y, Lu Y and Wei D. **2004**. Antioxidant activity of flavonoid-rich extract of *Hypericum perforatum* L *in vitro*. *Journal of Agriculture and Food Chemistry*, 52, 5032-5039.